## Estimating the Null Distribution

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#### Announcements

- HW 9: Due April 23 at 5:00pm, email TA Scott Liang at ricestat533@gmail.com
- Lectures: Today, April 23
- Take home exam
  - Assigned on Thursday April 23
  - Due April 29 at 5:00pm
  - Similar structure to Exams 1 and 2
  - Same policies as Exam 2
  - Strong focus on content in Efron, last third of course
- Lecture Format
  - Slides (plots / analyses in R)
  - .pdf and .R available on course website



Issues with Theoretical Null

Why Null Model May Fail

Modeling the Null Distribution with Normal



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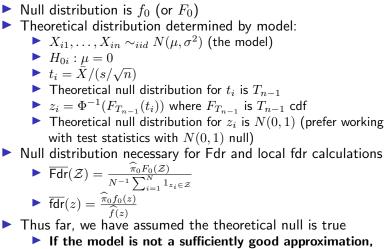
## Two Group Model

$$f_0(z)$$
 if  $y_i = 0$  (i.e.  $H_{0i}$  is true)  
 $f_1(z)$  if  $y_i = 1$  (i.e.  $H_{1i}$  is true)

• The marginal distribution of  $z_i$  is

$$f(z) = \pi_0 f_0(z) + \pi_1 f_1(z)$$

## Theoretical Null Distribution



then the inferences drawn from  $\overline{\rm Fdr}(\mathcal{Z})$  and  $\overline{\rm fdr}(z)$  may be misleading

## Leukemia Data

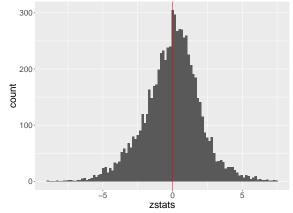
- 72 patients with leukemia
  - 47 with ALL (acute lymphoblastic leukemia)
  - 25 with AML (acute myeloid leukemia)
- $X_{ij}$  is expression of gene *i* for patient *j*
- Normalize expression values

$$x_{ij} = \Phi^{-1} \left( \frac{\operatorname{\mathsf{rank}}(X_{ij}) - 0.5}{N} \right)$$

where rank $(X_{ij})$  is rank of  $X_{ij}$  among  $X_{1j}, \ldots, X_{Nj}$ . Removes extreme outliers.

• Two sample t-tests on  $x_{ij}$  for  $i = 1, \ldots, 7128$ 

## z-statistic Distribution: Theoretical Null is Suspect



- Mean shifted positive
- Highly overdispersed relative to N(0,1)
- Two interpretations:
  - ▶ Many true alternatives, i.e.  $\pi_0 < 0.75$  (these do not have to follow null)
  - Theoretical N(0,1) is false

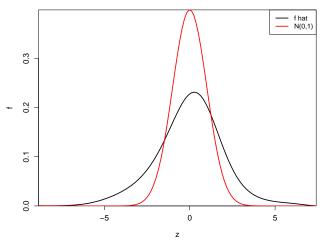
## Estimate $\pi_0$

#### • Estimate $\pi_0$ with Null Region Only Method

$$\widehat{\pi}_0 = \frac{N^{-1} \sum_{i=1}^N \mathbf{1}_{z_i \in \mathcal{A}_0}}{F_0(\mathcal{A}_0)}$$

► 
$$\mathcal{A}_0 = [\Phi^{-1}(0.25), \Phi^{-1}(0.75)] = [-0.67, 0.67]$$
  
► So  
 $\widehat{\pi}_0 = \frac{N^{-1} \sum_{i=1}^N \mathbf{1}_{z_i \in \mathcal{A}_0}}{0.5} \approx 0.593$ 

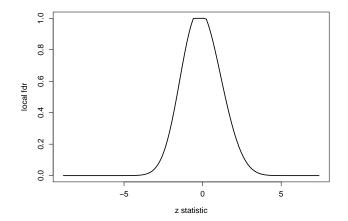
# Local fdr



Estimate f with Efron flexible exponential model.

$$\widehat{\mathsf{fdr}}(z) = \frac{\widehat{\pi}_0 \phi(z)}{\widehat{f}(z)}$$

## Local fdr



1539 genes with  $\widehat{\rm fdr} < 0.2$ 

## Summary of Results

- ▶  $\hat{\pi}_0 \approx 0.59$  suspiciously low (requires some knowledge about microarrays and cancer to make this determination)
- ▶ 1539 genes with  $\widehat{fdr} < 0.2$  is a lot
- Calculations quite sensitive to  $f_0$  (see more later)
- I get somewhat discrepant results from Efron here
  - Efron get  $\hat{\pi}_0 \approx 0.65$
  - Data may be slightly different, sample sizes do not match
  - I use mixture of my own code and Efron's locfdr package (on CRAN)
- ► Now: Discuss how / why theoretical f<sub>0</sub> may not be correct null distribution



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## Group Distributions

- Leukemia test statistics based on two-sample t-tests
- If AML and ALL not normally distributed, then test statistics may not follow t
  - Quality of normal approximation:
    - If both ALL and AML large, then test statistic is approx N(0,1) regardless of AML/ALL population distributions based on CLT
    - Transform original data to remove outliers
  - Challenges:
    - 25 AML cases, so CLT based normality may not hold
    - Difficult to find transformation that will work for all genes, recall there are 7128 t-tests
- For other models, checking assumptions / improving models even more difficult:
  - Example: Kidney cancer survival Computed test-statistics / p-values using Cox proportional hazards model. Assessing model assumptions / performing transformations with Cox is more difficult than two-sample t-tests

#### Independence

- Most test statistics assume n independent samples
- For example in two-group t-test  $x_{ij}$  are independent across j
- Possible causes of violation in Assumption
  - Matched pairs of patients collected in each group (age, sex, disease progression, etc.)
  - Samples processed in **batches**. For example x<sub>i1</sub>,..., x<sub>ik</sub> and x<sub>i.k+1</sub>,..., x<sub>i.n</sub> had expression measured on separate trays.
    - Many efforts to remove batch effects, but not always successful.
    - With very poor design, e.g. running all ALL patients in one batch and all AML patients on a second, can completely confound disease and batch effect.

## Covariates

- Often collect observational data
- ALL and AML samples not randomized units that were treated with ALL and AML
- Systematic differences in covariates between groups: age, sex, treatments, race, etc.
- Shifts in covariates may cause changes in expression, not related to disease type
- Example:
  - Gene i expression increases with age (all cells)
  - AML patients systematically older than ALL patients
  - Then gene i expression in AML tissue higher than in ALL tissue
  - But differential expression not caused by cancer
  - Healthy tissue from AML patients would have higher gene i expression than healthy tissue from ALL
- Tricky Issue: Standard null is wrong for gene i, but not wrong for the reason we care about. Issues related to causal inference.

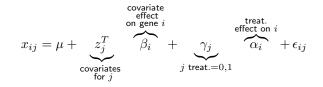
#### Fixing the Model

Best solution: Fix model for computing test statistics

- Use test statistics which do not require normality, e.g. Wilcoxon tests instead of t-tests
- Incorporate dependence, e.g. paired t-tests
- Use covariates in model fitting
  - Standard t-test (test statistic computed from α<sub>i</sub>):

$$x_{ij} = \mu + \underbrace{\gamma_j}_{j \text{ treat.}=0,1} \underbrace{\alpha_i}^{\text{treat.}} + \epsilon_{ij}$$

Model with covariates:



## Fixing the Model Not Always Feasible

- Only test statistics are available, not gene expression values or covariates
- Computation: More sophisticated models require more computation time
- Division of labor / scientific expertise: Determining appropriate normalization strategies / statistical models for gene expression may involve digging deeply into software pipelines and biological questions about which we have little expertise



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## Setup

Assumptions:

1.  $f_0$  follows a (possibly non-standard) normal:

$$f_0(z) \sim N(\delta_0, \sigma_0^2)$$

2.  $f_1(z) = 0$  for  $z \in \mathcal{A}_0$  (Null only region assumption)

**Goal:** Estimate  $\pi_0$ ,  $\delta_0$ ,  $\sigma_0^2$  and redo Fdr / local fdr calculations with estimated  $f_0$ .

**Note:** Assumptions are only approximations. May want to consider this as a sensitivity analysis exercise, i.e. How much do results change if consider non N(0, 1) null distribution?

## Estimation

#### Idea:

- Use only  $z_i$  in  $\mathcal{A}_0$  to estimate  $\delta_0, \sigma_0^2$ .
- Use fraction of data in  $\mathcal{A}_0$  to estimate  $\pi_0$

#### Notation:

Define

$$r_i = \begin{cases} 1 & z_i \in \mathcal{A}_0 \\ 0 & \text{o.w.} \end{cases}$$

$$r_i = \begin{cases} z_i & z_i \in \mathcal{A}_0 \end{cases}$$

$$z'_i = \begin{cases} -\infty & \text{o.w.} \end{cases}$$

• Data is 
$$(r_i, z'_i)$$
 pairs

 $\blacktriangleright$   $-\infty$  could be any value

• 
$$N_{\mathcal{A}_0} = \sum r_i$$
 (number of  $z_i$  in  $\mathcal{A}_0$ )

 $\blacktriangleright$   $z_i$  outside  $\mathcal{A}_0$  are "censored"

## Likelihood Function

$$p((r_i, z'_i)) = p(z'_i | r_i) p(r_i)$$

$$p(r_i)$$

$$r_i \sim Bern(\underbrace{\pi_0 H(\delta_0, \sigma_0^2)}_{\equiv \theta})$$

$$H(\delta_0, \sigma_0^2) = \int_{\mathcal{A}_0} \phi(z; \delta_0, \sigma_0^2) dz$$

$$p(z'_i | r_i)$$

$$Case r_i = 0:$$

$$p(z'_i | r_i = 0) = \begin{cases} 1 & z'_i = -\infty \\ 0 & o.w. \end{cases}$$

$$Case r_i = 1:$$

$$p(z'_i|r_i = 1) = \frac{\phi(z'_i; \delta_0, \sigma_0^2)}{H(\delta_0, \sigma_0^2)}$$

Assuming i.i.d. test statistics likelihood is:

$$\prod_{i=1}^{N} p((r_i, z'_i)) = \binom{N}{N_{\mathcal{A}_0}} \theta^{N_{\mathcal{A}_0}} (1-\theta)^{N-N_{\mathcal{A}_0}} \prod_{\{i: z_i \in \mathcal{A}_0\}} \frac{\phi(z'_i; \delta_0, \sigma_0^2)}{H(\delta_0, \sigma_0^2)}$$

## Notes on Likelihood

This is censored likelihood common in survival analysis:

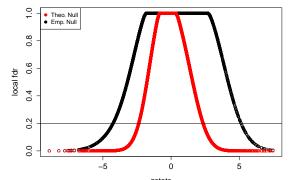
- $\triangleright$   $z_i$  outside  $\mathcal{A}_0$  are treated as censored.
- Replaced with placeholder  $-\infty$
- r<sub>i</sub> is indicator for censoring (1=not censored)
- Log likelihood is concave so MLE is unique
- Could attempt to use original data, e.g. in generalization of BUM / Patra-Sen mixture model:

$$F(z) = \pi_0 \Phi\left(\frac{z-\delta_0}{\sigma_0}\right) + (1-\pi_0)F_1(z)$$

where  $\Phi$  is N(0,1) and parameters are  $\sigma_0, \delta_0, F_1$ 

Disadvantage: Requires parametric assumptions on F<sub>1</sub> or complex procedures to non-parametrically estimate F<sub>1</sub>.

#### Application to Leukemia Data Estimates: $\hat{\delta}_0 = 0.136$ , $\hat{\sigma}_0 = 1.586$ , $\hat{\pi}_0 = 0.915$



Number of hypotheses with local  $\widehat{\text{fdr}} < 0.2$ :

- Theoretical null: 1501
- Empirical null: 244

#### Estimating null from data makes a huge difference!

## Application to Leukemia Data

Permutation tests can be used to approximate null distribution

- Randomly permute AML / ALL labels.
- Compute test statistics.
- Use resulting distribution as empirical null.
- Protects against t-statistic normality assumption.
- Does not have any effect for Leukemia data (permutation null distribution is approx N(0,1))
- Leukemia theoretical null violation likely due to hidden covariates.
  - Many genes show small levels of differential expression which are caused by covariates rather than AML/ALL.

## Summary / Preview

- Poor theoretical null can greatly change inferences
- Modeling  $f_0$  requires assumptions:
  - $f_0$  belongs to normal family, Null Only Region near 0
  - Rather than believing the results, can consider modeling f<sub>0</sub> as a type of sensitivity analysis
- Efron suggests that covariates may be biggest problem in causing theoretical null not to hold
- On Thursday Chapter 7: Estimation accuracy of fdr under dependence

